

Invertebrate Monitoring in Wetlands

Joel Chirhart Minnesota Pollution Control Agency Biological Monitoring Unit



What effects the quality of the wetlands we are sampling?



The Five Major Factors Which Determine the Integrity of Wetlands





What is so great about biological data?





The Big Picture





Why Macroinvertebrates?

- Invertebrates are commonly and widely distributed in many types of wetlands
- Invertebrates respond with a range of sensitivities to many kinds of pollution
- Many aquatic invertebrates complete their life cycles in wetlands, so they are exposed directly to the physical, chemical, and biological conditions within the wetland
- Aquatic invertebrates are important in wetland food webs for wildlife



Evaluating the health of a wetland requires a measure that integrates multiple factors



Economists rely on an integrative tool to assess economic condition

Index of Leading Economic Indicators

 Length of work week, unemployment claims, new manufacturing orders, vendor performance, net business formation, equipment orders, building permits, change in inventories, stock prices, and money supply



Doctors use multiple measures to assess human health

 Urine chemistry, blood-cell count, blood chemistry, blood pressure, heart-rate, cholesterol levels, body temperature, throat culture, weight, chest x-ray.



The Index of Biological Integrity (IBI)

- Biologist have developed the IBI as a multiple factor measure to assess the health of streams and wetlands.
- The factors (metrics) that comprise the IBI are measures of different components of the biological community that have been selected based on their ability to reflect human induced changes.



IBI Metrics

- Taxonomic Diversity (biodiversity)
- Number of Intolerant Groups
- Percentage of Tolerant Groups
- Percentage of Dominant Groups
- Trophic Structure (feeding behavior)
- Individual Health



Leech Taxa Metric – Number of kinds of leeches





 Corixidae Proportion Metric – Ratio of water boatman, to other hemipterans and beetles in the bottle trap sample





Dragonfly-Damselfly
 Metric – Number of
 kinds of dragonflies
 and damselflies
 (odonata)





- ETSD Metric –
 Number of kinds of
 Mayflies and
 Caddisflies, and the
 presence of odonata
 and fingernail clams
- Snail Taxa Metric

 Number of kinds
 of snails





• **Total Taxa Metric** – Number of kinds of invertebrates



- Invertebrates are sampled in the month of June or early July
 - Samples are taken during this "index period" in order to ensure that the macroinvertebrates are at size that makes them easy to identify
 - This index period also ensures that the majority of the invertebrates collected spent their lives maturing in the wetland being sampled and did not fly in from another nearby wetland.



- Invertebrate samples are collected in the shallow, near-shore area not deeper than 3 feet.
- Bottle-traps samples and dip-net samples
 are collected in the same general area of the
 wetland



 If very little vegetation is present, sample close to shore and any vegetation that might present









• If there is a vegetated border around the wetland, sample throughout the vegetated zone, from near shore up to 1 meter deep









• If the wetland has dense vegetation throughout, try to find open pockets in which to sample in the near shore area.











How to sample?

Bottle Traps





• Dip-netting



Bottle Trap





Bottle Trap Sample - Placement

- Place 6 bottle traps in 3 pairs along shoreline.
- The members of each pair should be 3-6 feet apart.
- Pairs should be spaced 20 ft apart.
- At least one pair should be in very shallow water (1
 foot or less), the others
 should be in water 2-3 feet
 deep





Bottle Trap Sample - Placement

- Bottle traps are set out for 2 nights
- Bottle traps should be place in water horizontally with no air bubbles inside.
- Funnel should be snapped in securely, clamp should be tightned.
- The top of the bottle trap should be 3-5 inches below the water surface.



Bottle Trap Sample - Retrieval

- Collect each pair of bottle traps into one jar (3 jars total for 3 pairs of bottle traps)
- Turn the bottle trap in the water the opening is facing upwards
- Raise the trap up dowel, remove the funnel, and pour the contents of the trap through a sieve.





Bottle Trap Sample - Retrieval

- Dislodge any critters stuck on the **inside** of the trap.
- Collect the second trap and pour into sieve.
- Flush the sieve a sample jar with 95% alcohol.
- If the sample takes up more than a third of the sample jar it should be split between two jars.
- Label the outside and the inside of the jar, using a media that resists alcohol (india ink or pencil)



Dip netting Tray





Dipnet Sample - Collection

- Each dipnet sample consist of two dipnetting efforts
- Dipnet in the near shore area in water up to one meter.
- Sample close to the edge and into vegetation.
- Using strong strokes, sweep the dipnet through the vegetation towards your body 3-5 times or until the net is full of vegetation









Dipnet Sample - Collection

- Empty the contents of the net on the hardware cloth screen.
- Spread the vegetation out, and pour some filtered site water over it.
- Spread the vegetation
 and repeatedely loosen it
 and tease through it for
 10 minutes. Remove the
 vegetation and repeat.









Dipnet Sample - Preservation

- After the second dipnetting process, empty the contents of the trays into a sieve.
- Backflush the sieve into a sample jar with 95% alcohol.
- If the sample takes up more than a third of the sample jar it should be split between two jars.
- Label the outside and the inside of the jar, using a media that resists alcohol (india ink or pencil)



Urban Wetlands: Not always impaired

Mud Lake, Plymouth

72Invert IBI30(Support)Non-Support)50Plant IBI18(Support)(Non-Support)



Agricultural wetlands: Not always impaired



70Invert IBI30(Support)(Non-Support)51Plant IBI18(Support)(Non-Support)